

Fig. 8 Neohendersonia kickxii (CBS 112403). a Conidioma on Fagus sylvatica. b Conidiogenous cell in vivo. c Conidia in vivo showing the paler basal cell. d Conidiogenous cells in vitro (arrows). e Conidia

sylvatica in Belgium. Sutton and Pollack (1974) subsequently combined Stilbospora kickxii into Neohendersonia, and N. kickxii was proposed as the type species, being the older name. After the inclusion of N. congoensis, the generic concept was expanded to include species with euseptate conidia (Sutton 1975). Recently, a third species, N. fagi, has been described from the same source in Italy (Wijayawardene et al. 2016). Currently, from the three species accepted in the genus, only N. kickxii has living type material, and, therefore, we cannot elucidate the monophyly of the genus and the species boundaries. However, the three species can be morphologically distinguished by the presence of euseptate conidia in N. congoensis and the size of the distoseptate conidia in N. fagi (30-48 \times 10-15 µm) and *N. kickxii* (21–45.5 × 12.5–20 μm) (Sutton 1975, 1980; Wijayawardene et al. 2016).

From the material examined here, the culture CPC 24865 was found to be phylogenetically distinct from the other isolates of *N. kickxii* and it could represent a different taxon. However, we prefer to not assign a name to this culture yet, since we did not find sufficient morphological characteristics to propose it as a different species.

Neohendersonia kickxii has been reported as a specific endophyte of beech twigs in Europe (Danti et al. 2002; Sieber 2007) and from beech bark in North American forests (Griesmer-Zakhar 2013), while *N. congoensis* and *N. fagi*

in vitro. **f** Second kind of conidia growing directly from the hyphae. Scale bars: $\mathbf{b}-\mathbf{e} = 10 \ \mu\text{m}, \mathbf{f} = 5 \ \mu\text{m}$

have been described on stems of *Aloe* or *Agave* in Congo and branch of *Fagus* in Italy, respectively.

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Spumatoria Massee and E.S. Salmon, Ann. Bot. Lond. 15: 350 (1901).

Classification: Ophiostomataceae, Ophiostomatales, Sordariomycetes.

Current generic circumscription: Saprobic, coprophilous. Ascomata perithecial, single, gregarious, immersed, erumpent, loculus globose with one or seldom two necks per perithecium. Ascomatal neck centrally, terete, apically paler and conspicuously long fimbriate, basally dark brown to black, felty, textura prismatica. Peridium dark brown to black, slightly hard, rough, basally with red brown hyphae, textura angularis. Paraphyses longer than the asci. Asci 8-spored, clavate, apically rounded and with an ocular chamber, pedicel mostly short and not furcate, thick-walled, bitunicate. Ascospores obliquely biseriate, overlapped, 2(-3)-celled, clavate, straight, hyaline, thin-walled, smooth, coarsely guttulate, septa smooth and thin-walled. Asexual morph sporothrix-like (in culture). Conidiophores more or less differentiated, unbranched, cylindrical. Conidiogenous cells polyblastic, sympodial, terminal, denticulate. Conidia solitary, dacrioid, hyaline. Blastoconidia growing directly from undifferentiated hyphae, lateral, globose to subglobose.

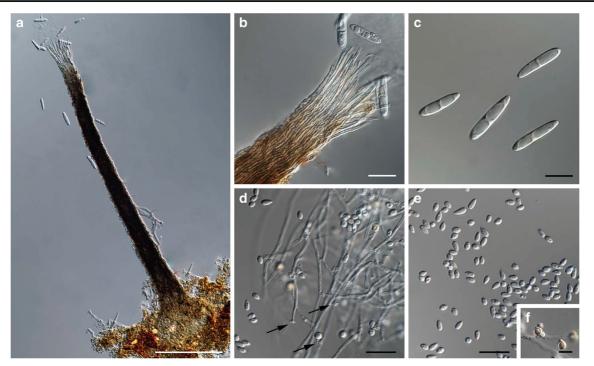


Fig. 9 Spumatoria longicollis (CBS 141464). **a** Ascoma. **b** Ostiolar hyphae. **c** Ascospores. **d** Conidiogenous cells (arrows denote the denticles). **e** Conidia. **f** Blastoconidia. Scale bars: $\mathbf{a} = 100 \ \mu\text{m}$, all others = 10 μm

Type species: *Spumatoria longicollis* Massee & E.S. Salmon 1901.

Spumatoria longicollis Massee and E.S. Salmon, Ann. Bot. Lond. 15: 351 (1901). Fig. 9

MycoBank: MB 171713

Description: Ascomata single, gregarious, immersed, erumpent, loculus globose with one or seldom two necks per perithecium, up to 500 μ m long \times 39–60 μ m wide. Ascomatal neck central, terete, apically paler and conspicuously long fimbriate, basally dark brown to black, felty, textura prismatica, up to 430 μ m long × 28–30 μ m wide. Peridium dark brown to black, slightly hard, rough, basally with red brown hyphae, textura angularis. Paraphyses longer than the asci. Asci dehiscent, 8-spored, clavate, apically rounded and with an ocular chamber, pedicel mostly short and not furcate, thick-walled, bitunicate. Ascospores obliquely biseriate, overlapped, 2(-3)-celled, clavate, straight, hyaline, thin-walled, smooth, coarsely guttulate, septa smooth and thin-walled, $(17)18-23(26) \times 4-7.5$ µm. As exual morph sporothrix-like (in culture). Conidiophores poorly differentiated, unbranched, cylindrical, up to 30 µm long. Conidiogenous cells polyblastic, sympodial, terminal, denticulate. Conidia dacrioid, hyaline, smooth- and thin-walled, $3-4.5 \times 1.5-$ 2 µm. Blastoconidia growing directly from undifferentiated hyphae, lateral, globose to subglobose, smooth- and thickwalled, light brown, $3-4 \times 2-4 \mu m$ wide.

Culture characteristics: Colonies on OA attaining 13– 20 mm diam. after 1 week at 25 °C, flat, scarce aerial mycelium, dark brown. On MEA flat, creamy with scarce aerial mycelium, dirty white turning dark brown with age.

Specimens examined: England, Essex, Epping Forest, horse dung, G.E. Massee & E.S. Salmon [Massee G.E and E.S. Salmon, Ann. Bot. Lond. 15, Fig. 27] (Lectotype designed here, MycoBank MBT373089). Netherlands, near Oostvoorne, dune grassland, on cow (Galloway) dung, 26 Jan. 2016, coll. J. van der Lee, det. R.K. Schumacher, iso. A. Giraldo (epitype designated here, CBS H-22665, MycoBank MBT373090; culture ex-epitype CBS 141464 = CPC 30521).

Notes: As far as we know, this is the first report of this fungus after its original description. The sexual morph was only observed in natural substrata; many attempts to obtain this morph in culture resulted only in the sporothrix-like asexual morph. Several macroscopic and microscopic pictures were observed to corroborate the identity from the specimen. In the original protologue, Massee and Salmon (1901) mentioned cylindrical asci, $110-113 \times 13-15$ µm, versus narrow clavate in the illustration, and 2-celled, ellipsoidal ('oblongis utrinque rotundatis') spores, $15-19 \times 5 \mu m$. Unfortunately, due to the dehiscent nature of the asci and the scarce material available, these structures were not observed in this study. The authors refer the perithecia at the conidium-bearing stage to the genus Rhynchophoma, a similar genus described from the wood of Tilia ulmifolia (Karsten 1884). However, the type of this fungus is apparently lost and, according to De Beer et al. (2013), this genus does not belong to either Ophiostomatales or Microascales.

Among the coprophilous fungi, Kathistes is one of the genera morphologically similar to Spumatoria in having transversely septate ascospores and perithecia with long and dark necks, but can be differentiated by its narrower ascospores, particular sporidiomata and absence of paraphyses (Malloch and Blackwell 1990). Based on their morphological similarities and common habit, they were placed together with Klasterskya in the Kathistaceae within Ophiostomatales (Hawksworth et al. 1995; Kirk et al. 2008). However, the analysis of the SSU region showed the affinity of this family to be with the Dothideomycetes rather than the Ophiostomatales (Blackwell and Spatafora 1994); consequently, Klasterskya and Spumatoria were excluded from the latter order based on the morphological characters as reported in the original description (De Beer et al. 2013). It is important to mention that, from the approximately 300 genera currently included in the Ophiostomatales, Spumatoria is the only genus showing septate ascospores. However, our results of the molecular data, which include LSU (Fig. 4), ITS and β tubulin (data not shown) loci, demonstrate the inclusion of the latter genus within the Ophiostomatales. Based on phylogenetic placement, Spumatoria could be considered a synonym of the older name Sporothrix (Hektoen and Perkins 1900). Although S. longicollis also have a sporothrix-like asexual state, it differs from other Sporothrix spp. based on septate ascospores, light coloured ascomata and dung-inhabiting biology. We, thus, suggest that further study including sequences of multiple gene regions and more taxa is needed in order to elucidate its relation with the type species of Sporothrix, S. schenckii and other species in the genus.

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References

- Ahmed SA, van de Sande WWJ, Stevens DA et al (2014) Revision of agents of black-grain eumycetoma in the order Pleosporales. Persoonia 33:141–154
- Aveskamp MM, de Gruyter J, Woudenberg JH, Verkley GJ, Crous PW (2010) Highlights of the Didymellaceae: a polyphasic approach to characterise Phoma and related pleosporalean genera. Stud Mycol 65:1–60

- Batzer JC, Arias MM, Harrington TC et al (2008) Four species of *Zygophiala* (Schizothyriaceae, Capnodiales) are associated with the sooty blotch and flyspeck complex on apple. Mycologia 100: 246–258
- Berkeley MJ, Broome CE (1875) Enumeration of the fungi of Ceylon. Part II. Bot J Linn Soc 14:29–141
- Blackwell M, Spatafora JW (1994) Molecular data sets and broad taxon sampling in detecting morphological convergence. In: Hawksworth DL (ed) First international workshop on Ascomycetes systematics. Plenum Press, New York, pp 243–248
- Boehm EW, Schoch CL, Spatafora JW (2009) On the evolution of the Hysteriaceae and Mytilinidiaceae (Pleosporomycetidae, Dothideomycetes, Ascomycota) using four nuclear genes. Mycol Res 113:461–479
- Campbell J, Ferrer A, Raja HA, Sivichai S, Shearer CA (2007) Phylogenetic relationships among taxa in the Jahnulales inferred from 18S and 28S nuclear ribosomal DNA sequences. Can J Bot 85:873–882
- Castlebury LA, Rossman AY, Jaklitsch WJ, Vasilyeva LN (2002) A preliminary overview of the Diaporthales based on large subunit nuclear ribosomal DNA sequences. Mycologia 94:1017–1031
- Cheewangkoon R, Crous PW, Hyde KD, Groenewald JZ, To-anan C (2008) Species of *Mycosphaerella* and related anamorphs on *Eucalyptus* leaves from Thailand. Persoonia 21:77–91
- Cheewangkoon R, Groenewald JZ, Summerell BA et al (2009) Myrtaceae, a cache of fungal biodiversity. Persoonia 23:55–85
- Cheewangkoon R, Groenewald JZ, Hyde KD, To-anun C, Crous PW (2012) Chocolate spot disease of *Eucalyptus*. Mycol Prog 11:61–69
- Chen Q, Jiang JR, Zhang GZ, Cai L, Crous PW (2015) Resolving the *Phoma* enigma. Stud Mycol 82:137–217
- Crous PW, Groenewald JZ (2011) Why everlastings don't last. Persoonia 26:70–84
- Crous PW, Groenewald JZ (2013) A phylogenetic re-evaluation of *Arthrinium*. IMA Fungus 4:133–154
- Crous PW, Wingfield MJ, Park RF (1991) Mycosphaerella nubilosa, a synonym of M. molleriana. Mycol Res 95:628–632
- Crous PW, Gams W, Stalpers JA, Robert V, Stegehuis G (2004) MycoBank: an online initiative to launch mycology into the 21st century. Stud Mycol 50:19–22
- Crous PW, Groenewald JZ, Wingfield MJ (2006a) *Heteroconium* eucalypti. Fungal Planet no. 10
- Crous PW, Slippers B, Wingfield MJ et al (2006b) Phylogenetic lineages in the Botryosphaeriaceae. Stud Mycol 55:235–253
- Crous PW, Braun U, Groenewald JZ (2007a) *Mycosphaerella* is polyphyletic. Stud Mycol 58:1–32
- Crous PW, Mohammed C, Glen M, Verkley GJM, Groenewald JZ (2007b) *Eucalyptus* microfungi known from culture. 3. *Eucasphaeria* and *Sympoventuria* genera nova, and new species of *Furcaspora*, *Harknessia*, *Heteroconium* and *Phacidiella*. Fungal Divers 25:19–36
- Crous PW, Schoch CL, Hyde KD et al (2009a) Phylogenetic lineages in the Capnodiales. Stud Mycol 64:17–47
- Crous PW, Summerell BA, Carnegie AJ et al (2009b) Unravelling Mycosphaerella: do you believe in genera? Persoonia 23:99–118
- Crous PW, Verkley GJM, Groenewald JZ, Samson RA (2009c) Fungal biodiversity. CBS Laboratory Manual Series no.1. CBS-KNAW Fungal Biodiversity Centre, Utrecht, the Netherlands
- Crous PW, Groenewald JZ, Shivas RG et al (2011) Fungal planet description sheets: 69–91. Persoonia 26:108–156
- Crous PW, Summerell BA, Shivas RG et al (2012) Fungal planet description sheets: 107–127. Persoonia 28:138–182
- Crous PW, Wingfield MJ, Guarro J et al (2013) Fungal planet description sheets: 154–213. Higher order classification of taxonomic novelties. Persoonia 31:186–296